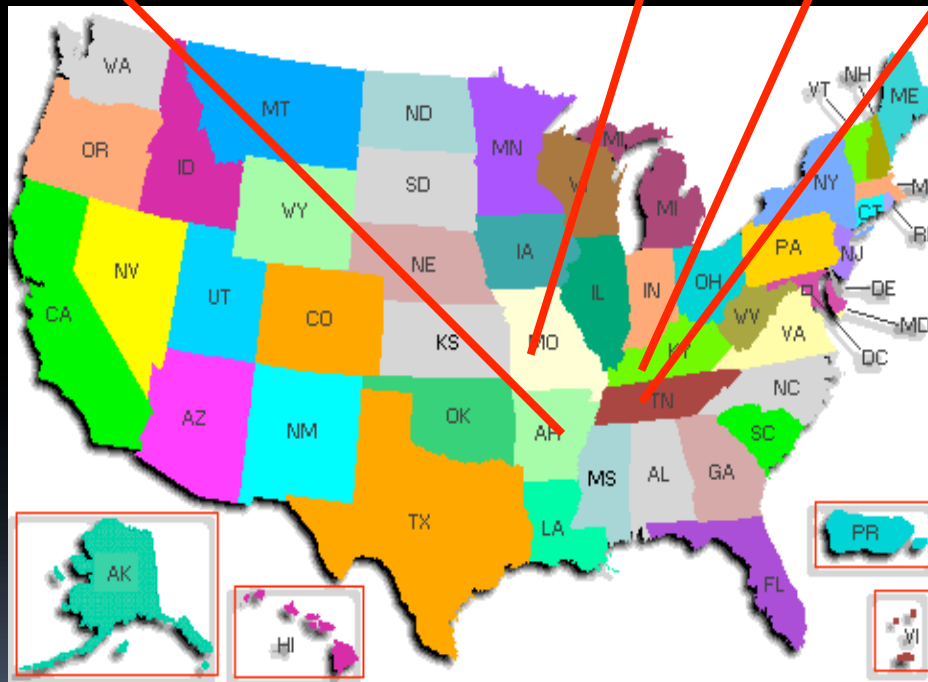
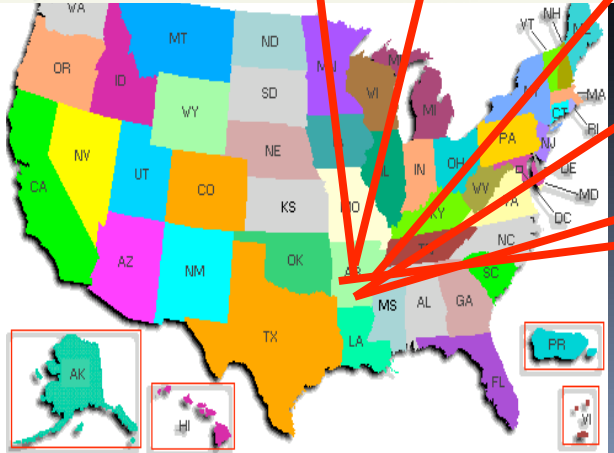


1971 - Arkansas

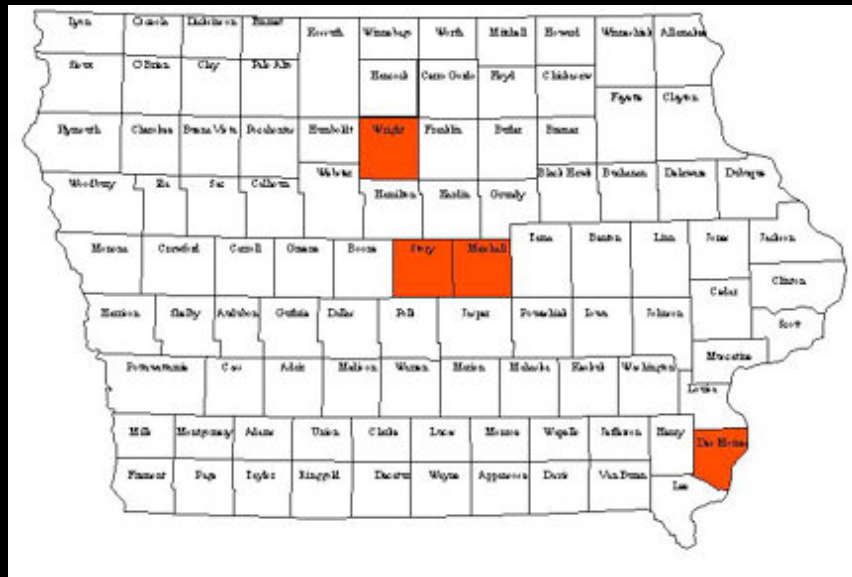
1984 - Missouri, Kentucky, Tennessee



Iowa State University

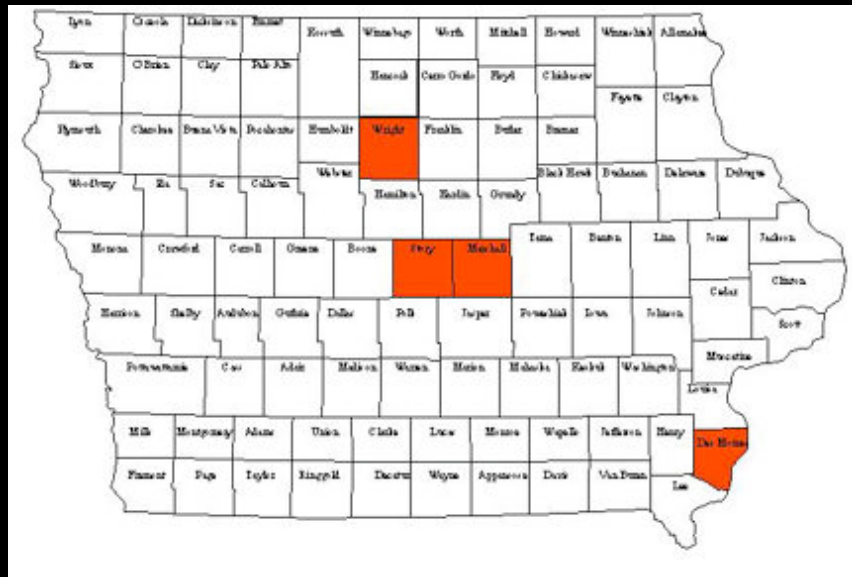


Iowa State University

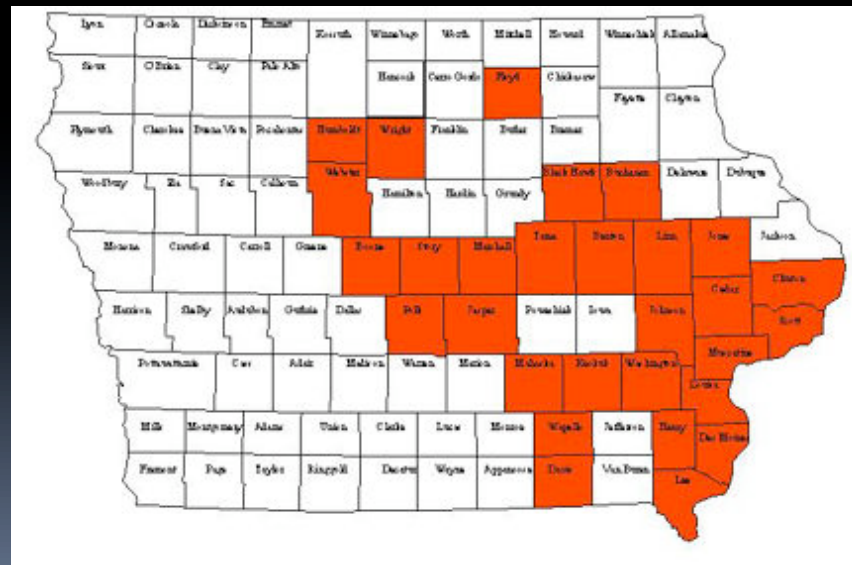


1994 SDS in Iowa

Iowa State University

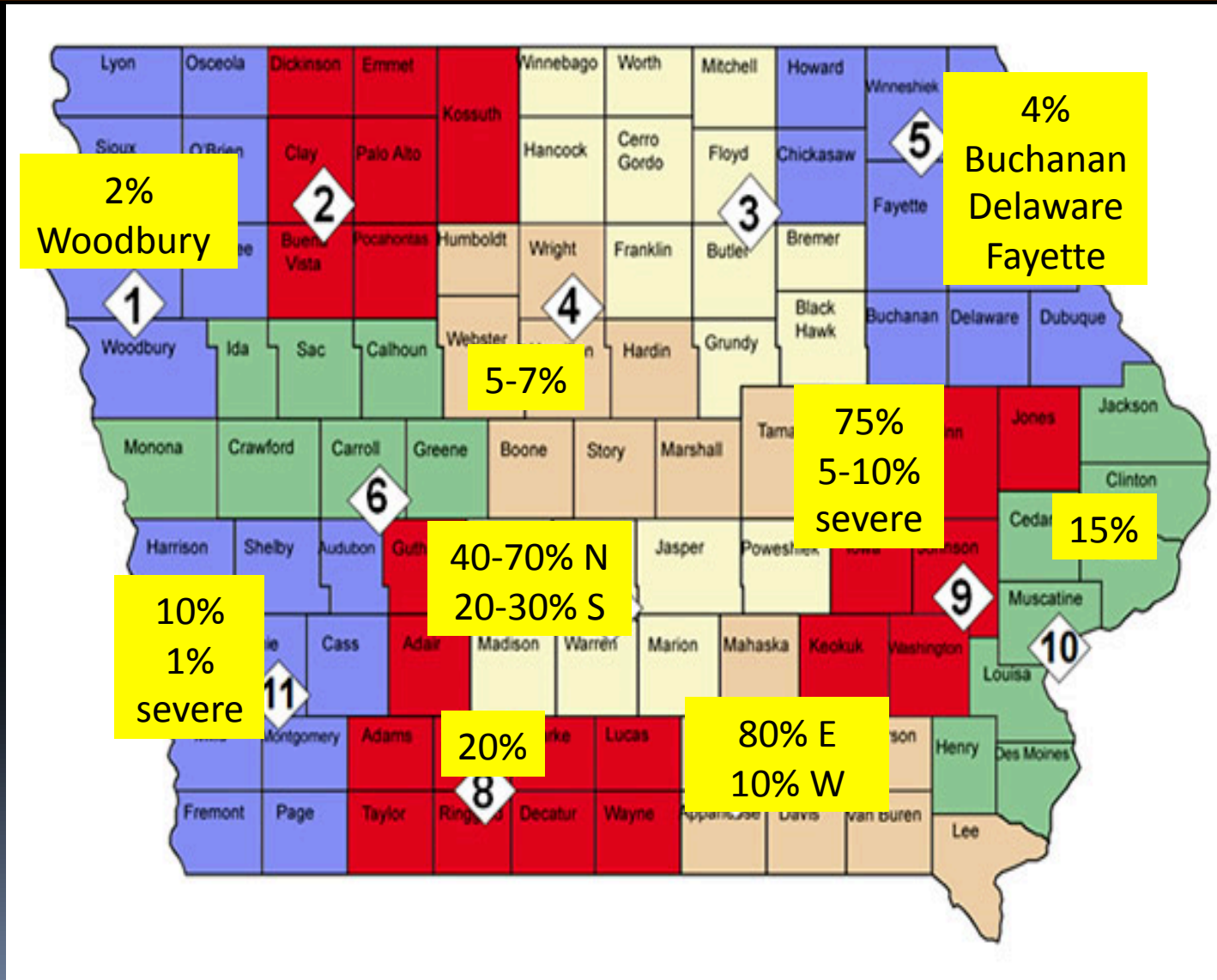


1994 SDS in Iowa



1998 SDS in Iowa

SDS Estimates in 2009



SDS – Fungal disease caused by *Fusarium virguliforme*

BREEDING EFFORTS TO IMPROVE SDS RESISTANCE IN SOYBEAN

Rationale for Breeding efforts

Relate to the plant

- Rapid spread of disease in soybean production areas
- Limited
 - number of genetic sources of resistance
 - knowledge of inheritance of resistance

Relate to the pathogen

- Screening tools amenable to differentiate small phenotypic effects
- Detailed genetic work to facilitate breeding resistance work in soybean

Breeding efforts

- Several groups are developing populations by crossing R x S genotypes
- Some are using screening data and molecular information to select for resistance
- At ISU, we still cross R x S, and also starting to develop populations by crossing R x R genotypes

Sources of SDS resistance

Breeding at ISU

- LS98-0582
- LS99-2235
- LS94-3207
- Ripley

Sources of SDS resistance
Breeding at ISU

LS98-0582

LS99-2235

LS94-3207

□ Ripley

	Disease Index				
	2004				
	IA	IA	Can	IL	IL
AR10SDS	1	9	1	1	16
MN 1606 SP (Res)	2	0	1	0	8
Venus RR (Res)	0	0	0	15	4
5171 RR (Susc)	4	8	0	15	20
	2005				
AR10SDS	0		14	0	0
MN 1606 SP (Res)	0		26	0	0
Venus (Res)	0		15	0	0
5171 RR (Susc)	0		19	2	20
	2007				
AR10SDS		2 (MN)	1 (IL)	0	0
MN1606 SP (Res)		10	0	0	0
Venus (Res)					
5171 RR (Susc)		20	8	4	17

The molecular information available

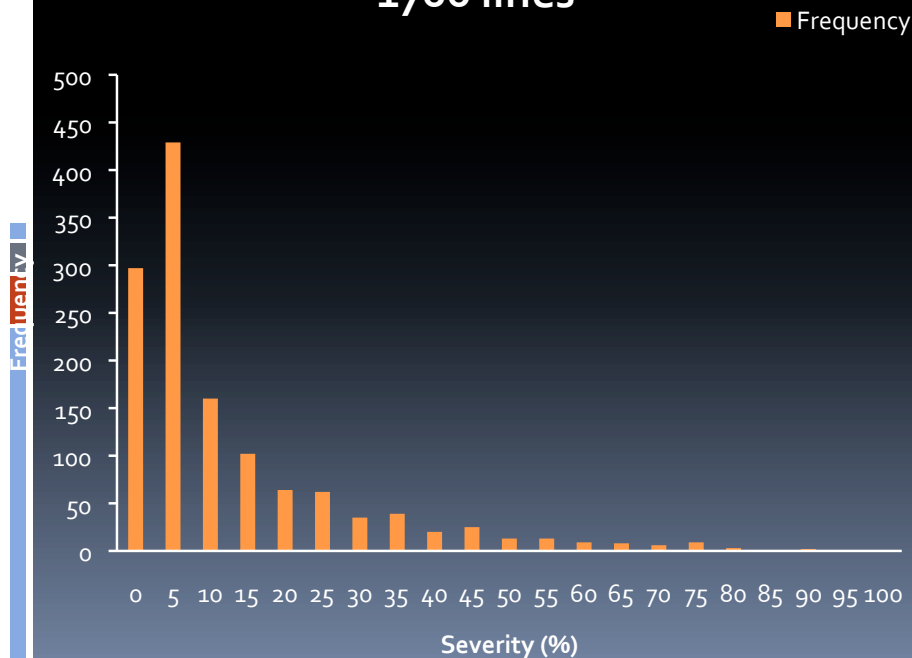
Population	# of QTL	Chromosomes
Essex x Forrest	9	13, 16, 6, 20, 13
Pyramid x Douglas	3	3,6,18
Hartwig x Flyer	2	18,8
Ripley x Spencer	3	4,17,19
PI567374 x Omaha	2	17, 20

Simultaneous work on

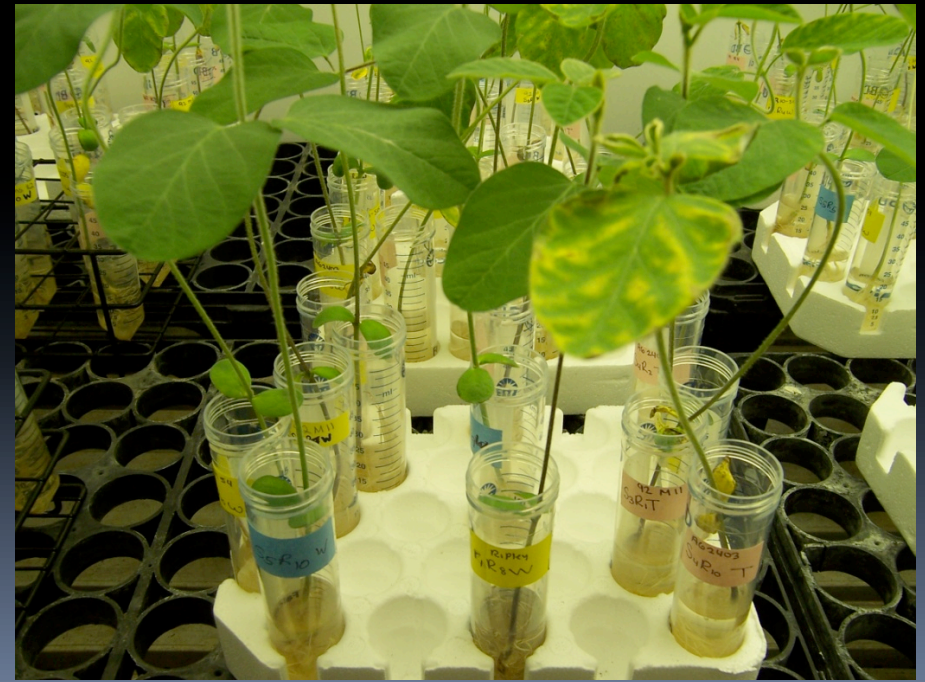
- I. Breeding for germplasm releases
 - Germplasm release - spring 2010
 - Searching for the proper combination of yield, SDS and MG for Iowa
- II. Considering interactions
 - SDS and SCN
 - SDS and BSR
- III. Research
 - modifications to the screening method (Luckew & Leandro)
 - new screening methods - toxin use (Bhattacharyya's lab)
 - molecular marker development
 - search for new genes

- Find the number of QTLs that will confer adequate resistance
- Screen ~1700 lines for differing SDS resistance
 - Using the classic screening method

Distribution of Severity among the ~1700 lines



- **The second screening method**
 - **Toxin assay using cultured filtrate**
 - **Mont-1 isolate**
 - **Plants with intact roots immersed in diluted toxin**
 - **Ten genotypes**



Details of Inbred Soybean Lines

Population Name	Susceptible parent	Resistant parent	No. of Populations	No. of Subpopulations
AX19286	A95-684043	LS94-3207	11	391
AX19287	A95-684043	LS98-0582	10	435
AX19288	A95-684043	LS99-2235	1	392
AX19289	IA1006	LS94-3207	4	446
AX19290	IA1006	LS98-0582	3	471
AX19291	IA1006	LS99-2235	1	85
AX19294	IA2050	LS94-3207	8	461

Screening of AX19286 subpopulations

2 slight chlorotic flecks/ blotches



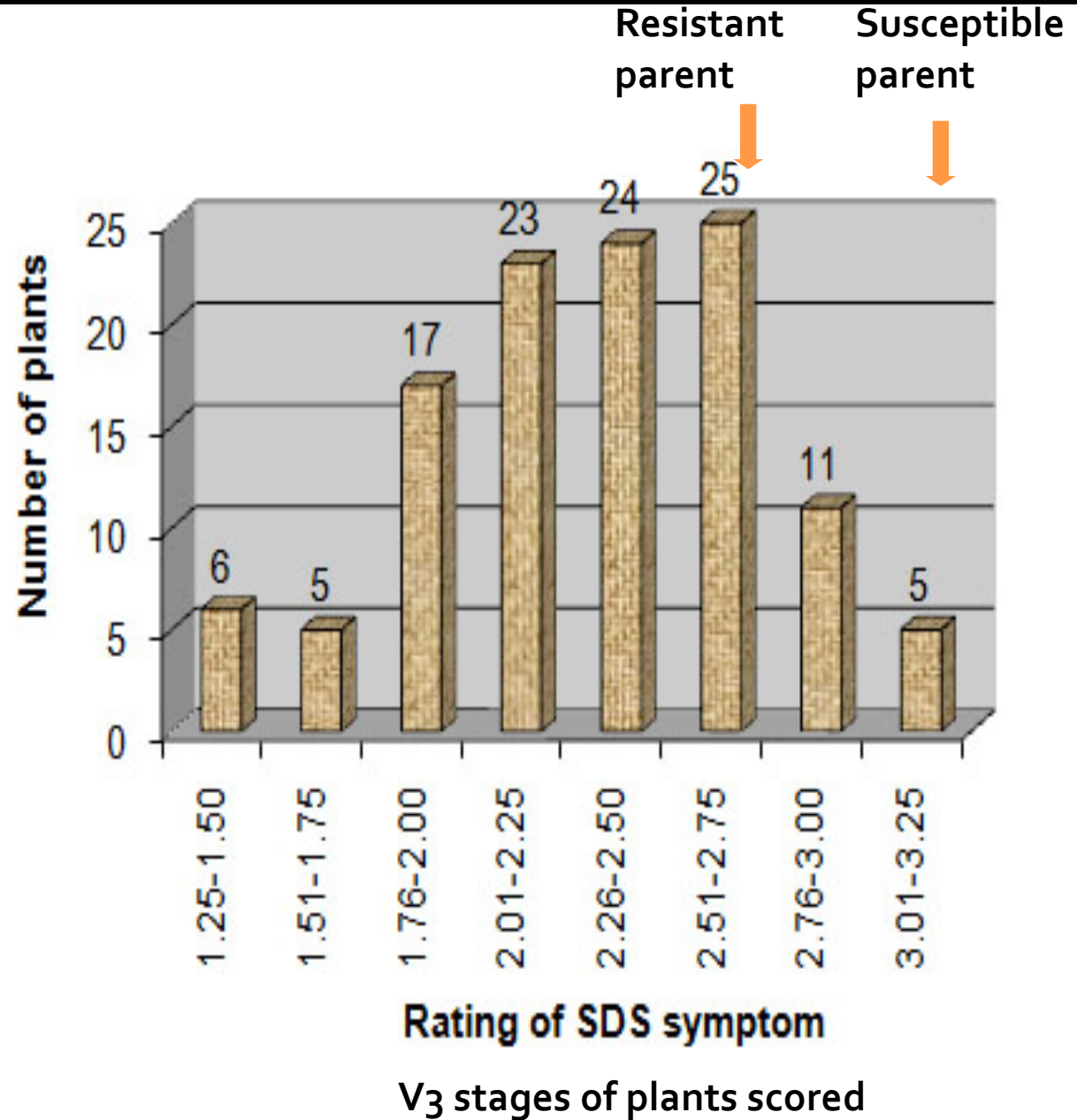
3 Leaf with inter-veinal chlorosis



4 - necrosis + chlorosis



5 extended Necrosis + cupping of leaves



Therefore,

- The work is undergoing in several breeding labs, with interesting results
- Important work is underway on the pathogen
- Screening techniques are modified and new ones are under development, serving phenotyping efforts, better and more efficiently
- Lines in early maturity groups are and will be available for breeding for disease resistance.
- And, the collective work continuous